RESEARCH REPORT

Application of phage display for the development of a novel inhibitor of PLA_2 activity in Western cottonmouth venom

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ABSTRACT

Snakebite envenomation is an important global health concern. The current standard treatment approach for snakebite envenomation relies on antibody-based antisera, which are expensive, not universally available, and can lead to adverse physiological effects. Phage display techniques offer a powerful tool for the selection of phage-expressed peptides, which can bind with high specificity and affinity towards venom components. In this research, the amino acid sequences of Phospholipase A_2 (PLA_2) from multiple cottonmouth species were analyzed, and a consensus peptide synthesized. Three phage display libraries were panned against this consensus peptide, crosslinked to capillary tubes, followed by a modified surface panning procedure. This high throughput selection method identified four phage clones with anti-PLA_2 activity against Western cottonmouth venom, and the amino acid sequences of the displayed peptides were identified. This is the first report identifying short peptide sequences capable of inhibiting PLA_2 activity of Western cottonmouth venom *in vitro*, using a phage display technique. Additionally, this report utilizes synthetic panning targets, designed using venom proteomic data, to mimic epitope regions. M13 phages displaying circular 7-mer or linear 12-mer peptides with antivenom activity may offer a novel alternative to traditional antibody-based therapy.

KEYWORDS: Antivenom, phage display, M13 phage, phospholipase A_2, *Agkistrodon piscivorus* species, *in silico* design, Western cottonmouth

INTRODUCTION

Snakebite envenomation is a global health concern and classified as a neglected tropical disease by the World Health Organization (WHO 2007, 2010; Chippaux, 2017). Approximately 5.4 million people are victims of snakebite worldwide each year, with 81,000-138,000 deaths and over 400,000 suffering from long-term disability, constant wound care, amputations and ongoing psychological morbidity. Nearly 9,000 cases of snakebite envenomation are reported in the United States and Canada every year (Smith et al, 2014; Mowry et al, 2013).

Venoms are extremely complex, consisting of hundreds of proteins, peptides, enzymes and non-enzymatic toxins (Hall et al, 2016). Through advancements in whole genome studies, venomics has evolved to assess toxin composition directly and indirectly via proteome- or transcriptome-based methodologies. This research has shown that venom composition varies between and within species due to ontogenetic and geographical variability, and this complexity of venom has led to inconsistent antivenom potency. For example, the same antivenom produced in different geographic regions of India exhibits different therapeutic potential, indicating that the current antivenom manufacturing techniques do not adequately account for the diverse venom composition (Warrell et al, 2013). Restricted availability of species-specific antivenoms can be circumvented by designing and producing high potency multipurpose antivenoms effective against multiple species of snakes (Calvete et al, 2009). Antibody-based antivenoms are routinely prepared from the plasma of host animals immunized with venoms from multiple snake species. While this is a successful therapeutic approach for some snake species, there remain limitations in safety, efficacy, and economic aspects of manufacturing. Immediate

66 hypersensitivity reactions following administration of Crotalidae polyvalent immune Fab
67 antivenom, including urticaria, dyspnea, hypotension, tachycardia, chest pain, and nausea may
68 occur in 19% of patients, and the range of serum sickness incidence including fever, myalgia,
69 epigastric pressure, and arthralgia is 6% - 23% (Schaeffer et al, 2012). In the United States the
70 most typical antivenom therapy using Crotalidae polyvalent immune Fab could cost more than
71 $18,000 and its usage can be a financial burden for patients and hospitals (Weant et al, 2010).
72 Thus, there is a need for a novel approach to develop antivenom solutions that are capable of
73 rapid production and maximal efficacy without adverse reactions. Phage display techniques offer
74 a comprehensive tool for the selection of phage-expressed peptides which have affinity for
75 unique targets normally inaccessible to antibody-based methods (Wu et al, 2016). Hence, a
76 phage display technique using composite consensus targets based on venom proteomic data
77 could lead to the development of a broad spectrum inhibitor of multiple venoms.
78
79 To demonstrate proof-of-concept of phage display for the development of venom component
80 inhibitor, the venom of the Western cottonmouth, *Agkistrodon piscivorus leucostoma*, a
81 venomous New World pit viper found in North and Central America (Lomonte et al, 2014), was
82 selected. Western cottonmouths inhabit a large geographic area and are found along the coast of
83 the Gulf of Mexico into southeastern and central Texas, presenting a concern to civilian
84 populations, as well as military operational and training sites across this region. Major
85 components of the Western cottonmouth venom are phospholipases A_2 (PLA_2, 43.2%),
86 metalloproteases (26.1%) and serine proteases (10.1%) (Lomonte et al, 2014). The clinical
87 effects of envenomation are the initiation of the coagulation cascade and blockage of
88 neuromuscular transmission. Snake venom PLA_2 enzymes interfere in the normal physiological

89 processes of victim animals and induce a wide variety of pharmacological effects including
90 neurotoxicity (Harris and Scott-Davey 2013), myotoxicity (Teixeira et al, 2003), and
91 anticoagulant effects (Saikia et al, 2013).
92
93 In this study, conserved PLA_2 sequences from several species of cottonmouth snakes were
94 identified and mapped to the three-dimensional (3D) crystal structure of PLA_2 to determine their
95 solvent accessibility. Phage-displayed peptides with affinity towards the target consensus
96 sequence were tested for the generation of potential venom inhibitors.
97
98 **MATERIALS AND METHODS**
99
100 **Designing consensus sequence**
101 Queries against the UniProt database were used to gather amino acid sequences of target
102 peptides. Sequences were uploaded to Basic Local Alignment Search Tool (BLAST) to
103 determine homologous regions using default parameters. Consensus peptides were designed
104 based on areas of homology, and areas corresponding to active sites were given priority during
105 the design process. The original sequences were aligned to the consensus sequence to ensure
106 homology and identity. Once the validity of the consensus sequence was confirmed, the
107 sequence was overlaid onto a crystal structure of the target protein. Crystal structure information
108 was unavailable for Western cottonmouth PLA_2, thus Eastern cottonmouth PLA_2 (PBD: 1PPA)
109 was used to map consensus sequence. JSmol Viewer, an open-source Java viewer, from Research
110 Collaboratory for Structural Bioinformatics Protein Data Bank (Berman et al, 2002) was used to
111 visualize the 3D mapping of the solvent exposed regions of the target peptide.

Screening of peptides with affinity to the consensus sequence

A cleavable crosslinker, Sulfo-LC-SPDP (sulfosuccinimidyl 6-[3'-(2-pyridyldithio)propionamido]hexanoate) was used to bind the target peptide to the interior of an aminosilylated glass capillary tube. Briefly, a glass capillary tube was washed with acetone and treated with aminosilane reagent. The silylated glass capillary tube was then modified with 10 mM Sulfo-LC-SPDP. After one hour incubation, the crosslinked capillary tube was rinsed twice with Coupling Buffer (50mM Phosphate, 150mM NaCl, 10mM EDTA, pH 7.2). The Sulfo-LC-SPDP modified capillary tube was filled with 5 µl of the target PLA_2 consensus peptide (10mg/ml) and incubated overnight at 4^oC to complete the crosslinking reaction. Phages were panned against target peptide using a modified panning method (Fralick, Chadha-Mohanty, and Li 2008). Briefly, a Marprene 0.5mm bore 1.6mm wall tubing (Watson-Marlow, Inc., Wilmington, MA) circuit containing a 10µl capillary tube with crosslinked peptide was inserted into a peristaltic pump (Heidolph North America, Elk Grove Village, IL). The tubing circuit was blocked for one hour with Blocking Buffer [100mM $NaHCO_3$, (pH 8.6), 5mg/ml bovine serum albumin (BSA)], then rinsed six times with 1x TBST [TBS + 0.1% (v/v) Tween-20]. Phage libraries (PhD -7, -12, -C7C Phage Display Peptide Library Kits, New England Biolabs, Ipswich, MA) were diluted to a working concentration of $1x10^{11}$ PFU/ml in 1x TBST, added to the circuit, and slowly circulated at a speed of <5 rpm for one hour at room temperature (RT). The tubing circuit was rinsed ten times with 1x TBST to facilitate removal of non-binding phages. After siphoning off non-binding phages, phages with high affinity to the target peptide were eluted using either Elution Buffer (25mM Dithiothreitol, pH 8.5) at 37^oC for Ph.D.-7 and Ph.D.-12 libraries or Acidic Elution Buffer [200mM Glycine-HCl (pH 2.2), 1mg/ml BSA] at RT for

Ph.D.-C7C phage library. Eluted high-affinity phages were collected in Eppendorf tubes and the Ph.D.-C7C eluent was neutralized using 1M Tris-HCl (pH 9.1). The eluate was diluted 1:1000 in phosphate buffered saline (PBS), then serially diluted and titered out for individual plaques on soft agar plates. An aliquot of the primary panning eluate was amplified for a new population of M13 phages up to $1x10^{12-13}$ PFU/ml for secondary panning. A second round of panning was carried out with a more stringent wash step (5x TBST). Individual phage clones from the secondary panning elute were selected on soft agar plates and amplified as before up to $1x10^{12-13}$ PFU/ml. These clones were picked separately and grown up individually for PLA_2 inhibition assay. To monitor the accuracy of panning process, the polyclonal phage isolates from the primary panning were mixed with venom at varying concentrations ($3.2x10^{12}$ PFU/ml and $8x10^{11}$ PFU/ml) and their anti-PLA_2 activity was measured.

Phage harvesting and quantification

Phages were harvested and quantified according to the manufacturer's protocol. Briefly, phages were harvested using a polyethylene glycol precipitation method (Yu and Smith, 1996) and resuspended in PBS (pH 7.5). Collected phages were titered on isopropyl β-D-1-thiogalactopyranoside/5-bromo-4-chloro-3-indolyl β-D-galactopyranoside (50mM IPTG/40mM X-Gal) Luria-Bertani medium. Phage clones were individually picked from soft agar titer plates and amplified based on the manufacturer's protocol for further testing.

Venom

Snake venoms [Western cottonmouth, Eastern diamondback rattlesnake (*Crotalus adamanteus*), Western diamondback rattlesnake (*Crotalus atrox*), Mojave rattlesnake (*Crotalus scutulatus*),

158 and broad-banded copperhead (*Agkistrodon contortrix laticinctus*)], were obtained in lyophilized
159 form from the National Natural Toxins Research Center (Kingsville, TX). Venoms were
160 reconstituted at a concentration of 10mg/ml in PBS (pH 7.5) and diluted to working
161 concentrations.
162
163 **PLA_2 Activity Assay**
164 PLA_2 activity was measured using the EnzChek Phospholipase A_2 Assay kit (Invitrogen, Eugene,
165 OR) with modification. Briefly, the concentration of Western cottonmouth venom was optimized
166 to be 4.88 mg/ml for the linearity of fluorescence signal from the plate reader. The selected
167 phage clones were diluted to 1x 10^{12} PFU/ml and incubated with Western cottonmouth venom
168 for 30min at RT. A non-specific M13 phage clone was used as a negative control. After adding
169 reaction substrate to phage and venom mixture, fluorescence (excitation at 485nm and emission
170 at 528nm) was measured for 20min at one minute intervals to monitor the reaction. The overall
171 PLA_2 activity was defined by the fluorescence level at the 10min time point.
172
173 **Cross-species Inhibition**
174 The inhibitory effect of monoclonal phage isolates on venoms beyond Western cottonmouth was
175 determined using the EnzChek Phospholipase A_2 Assay kit with modification. Venom from the
176 Western cottonmouth, Eastern diamondback rattlesnake, Western diamondback rattlesnake,
177 Mojave rattlesnake, and broad-banded copperhead, was diluted to a working concentration of
178 4.88µg/ml. Diluted venom was incubated with 1x10^{12} PFU/ml phage clones for 30min at RT. A
179 non-specific M13 phage clone was used as a negative control. After adding reaction substrate to
180 phage and venom mixture, fluorescence (excitation at 485nm and emission at 528nm) was

measured for 20min at one minute intervals to monitor the reaction. The overall PLA_2 activity was defined by the fluorescence level at the 10min time point.

DNA Sequencing

The double-stranded DNA (dsDNA) replicative form (RF) isolates of monoclonal phage from single plaque-infected ER2738 cultures were extracted using the QIAprep Spin Miniprep Kit (Qiagen, Germantown, MD) following manufacturer's directions. Extracted dsDNA RFs were sequenced upstream of the peptide insertion using the -96 gIII sequencing primer supplied with the Ph.D. Phage Display Peptide Library Kit. DNA sequences were translated to amino acid sequences using the ExPASy Translate tool from the Swiss Institute of Bioinformatics.

Statistical Analysis

All experiments were conducted in triplicate and each fluorescence reading from the PLA_2 reaction mixture was measured from three separate wells. The means and standard deviations were calculated from relative fluorescence units. Student's *t*-test was used to determine the level of significance ($p < 0.05$).

RESULTS

The BLAST report identified a region of 120 amino acids with high consensus. A section of 57 amino acids annotated as the active (yellow highlight) and metal binding site (red characters) for PLA_2 (Figure 1a) is shown. The section corresponding to the active site was matched to the predicted crystal structure of PLA_2 (PDB: 1PPA) from Eastern cottonmouth venom, and the

surface-exposed residues were annotated (Figure 1b). The consensus peptide used as the panning target shares ≥95% homology with the major North American Crotalid (rattlesnakes, copperheads and cottonmouths) PLA_2 proteins. During the panning process, polyclonal phage isolates from the primary panning process showed PLA_2 inhibition activity in a concentration-dependent manner (Figure 2). Two different concentrations of polyclonal anti-PLA_2 isolates ($3.2x10^{12}$ PFU/ml and $8x10^{11}$ PFU/ml) were incubated with Western cottonmouth venom, inhibiting PLA_2 activity by 55% and 15% of uninhibited venom. Since phages from the primary panning showed inhibition of PLA_2 activity, the secondary panning was conducted with these eluted phages in more stringent conditions (5x TBST). After the second round of phage panning, the eluent was amplified and tested against Western cottonmouth venom again. Four individual phage clones were selected from polyclonal mixture based on their anti-PLA_2 activity (Figure 3). These clones inhibited 30% to 60% of PLA_2 activity from Western cottonmouth venom. Phage DNA sequencing from the selected monoclonal isolates identified four unique sequences and their corresponding peptide sequences (Table 1). Three of the sequenced isolates were from the Ph.D.-C7C library, showing three circular 7-mer motifs (Ph.D.-C7C-6, Ph.D.-C7C-7, and Ph.D.-C7C-9). The fourth was from the Ph.D.-12 library with a linear 12-mer peptide (Ph.D.-12-7). Cross-species anti-PLA_2 activity was tested against five major snake venoms in North America (Western cottonmouth, Eastern diamondback rattlesnake, Western diamondback rattlesnake, Mojave rattlesnake, and broad-banded copperhead) using one of selected anti-PLA_2 clones (Ph.D.-12-7). Results showed approximately 40% inhibition in Western cottonmouth venom and 30% inhibition in the other crotalid venoms (Figure 4).

DISCUSSION

227

228 A consensus peptide, based on sequence homologous of PLA_2 in cottonmouth species venom,
229 proved to be a viable target for the selection of anti-PLA_2 peptides displayed on M13 phages.
230 Utilizing venom proteomic data, epitope design could be optimized *in silico* to produce a
231 consensus antigenic determinant. This *in silico* approach could enhance the effective selection
232 from a phage display library by targeting areas closely related to the venom activity, in contrast
233 to serum-based methods that produce 5% - 36% therapeutically relevant antibodies (Laustsen et
234 al, 2017). Successful selection of anti-PLA_2 display phages which can inhibit a major component
235 of Western cottonmouth venom, PLA_2, could open the door to targeting other venom
236 components, including metalloprotease or serine protease, utilizing this approach. Selection of
237 antivenom phages was completed within four months, including design and synthesis of a novel
238 consensus peptide, panning several Ph.D. libraries, and characterizing inhibitory effects.
239

240 While the observed highest inhibitory rate was 60%, a synergistic inhibitory effect could be
241 expected after combining all the selected anti-PLA_2 phages. Because four unique sequences of
242 selected peptides displayed on M13 phages would target different regions of the PLA_2 active
243 sites, a more comprehensive inhibitory effect could be observed. Expansion of target venom
244 components is currently underway and the design of consensus peptides for both metalloprotease
245 and serine protease is being formulated. Once inhibitory phages for the three major venom
246 components have been identified, a phage cocktail will be assembled and its synergetic effect
247 will be tested against whole venom in an animal model to confirm the efficacy in practical
248 application. In addition to screening individual venom components in Western cottonmouth

venom, the universal consensus sequences of six other venoms from the Crotalidae subfamily have been identified for screening, incorporating features of all seven Crotalidae snake venoms.

Antivenom development and production towards the venom of small animals (eg. scorpion and spider) is hindered by the limited amounts of venom available for collection (Laustsen et al, 2016). In this study, only 5 µl of consensus venom peptide (10 mg/ml) was needed for the panning process. *In silico* design and synthesis of consensus peptides could provide an alternative solution to the limitation of venom availability. Design of *in silico* peptides could utilize transcriptome annotations to identify potential epitopes of unknown or punitive toxins. In addition, the design of consensus peptides could be based on shared homology across several genera or species. A future therapeutic product developed from multiple composite consensus peptides could show general effectiveness across venom components from several snake species in the geographic region. This design methodology could be expanded to include target molecules beyond proteins like carbohydrates and metals that are integral to venom components.

CONCLUSIONS

To the best of our knowledge, this is the first report identifying short peptide sequences expressed on M13 phages that can inhibit PLA_2 activity of Western cottonmouth venom *in vitro*. The phage display methods conducted in this study could be implemented for the development of novel antivenom. Using BLAST search tool and subsequent categorization of venom sequences, the consensus target peptides of multiple peptide families could be systematically selected and used for the development of a universal antivenom covering snakes, reptiles, arachnids, sea

jellies, or other venomous species. Future work is needed to investigate the synergetic effects of selected phages on target venom components.

DISCLAIMER

The views expressed in this article are those of the author and do not necessarily reflect the official policy or position of the Department of the Navy, Department of Defense, nor the United States Government. This work was supported and funded by the Naval Medical Research Center's In-house Laboratory Independent Research program using work unit number G1606. The authors are military service members or contract employees of the United States Government. This work was prepared as part of their official duties. Title 17 U.S.C. §105 provides that 'Copyright protection under this title is not available for any work of the United States Government.' Title 17 U.S.C. §101 defines a United States Government work as a work prepared by a military service member or employee of the United States Government as part of that person's official duties. Approved for public release; distribution unlimited.

STATEMENT OF COMPETING INTERESTS

One patent is pending on the selected phages with antivenom activity and their displayed peptides. No additional competing interests were declared by the authors.

LIST OF ABBREVIATIONS

295 3D: Three-dimensional

296 BLAST: Basic Local Alignment Search Tool

297 IPTG: Isopropyl β-D-1-thiogalactopyranoside

298 PBS: Phosphate buffered saline

299 PFU: Plaque-forming unit

300 PLA_2: Phospholipase A_2

301 RF: Replicative form

302 RT: Room temperature

303 Sulfo-LC-SPDP: Sulfosuccinimidyl 6-[3'-(2-pyridyldithio)propionamido]hexanoate

304 TBS: Tris-buffered saline

305 TBST: TBS with 0.1% (v/v) Tween-20

306 X-Gal: 5-bromo-4-chloro-3-indolyl β-D-galactopyranoside

307

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362

FIGURE LEGENDS

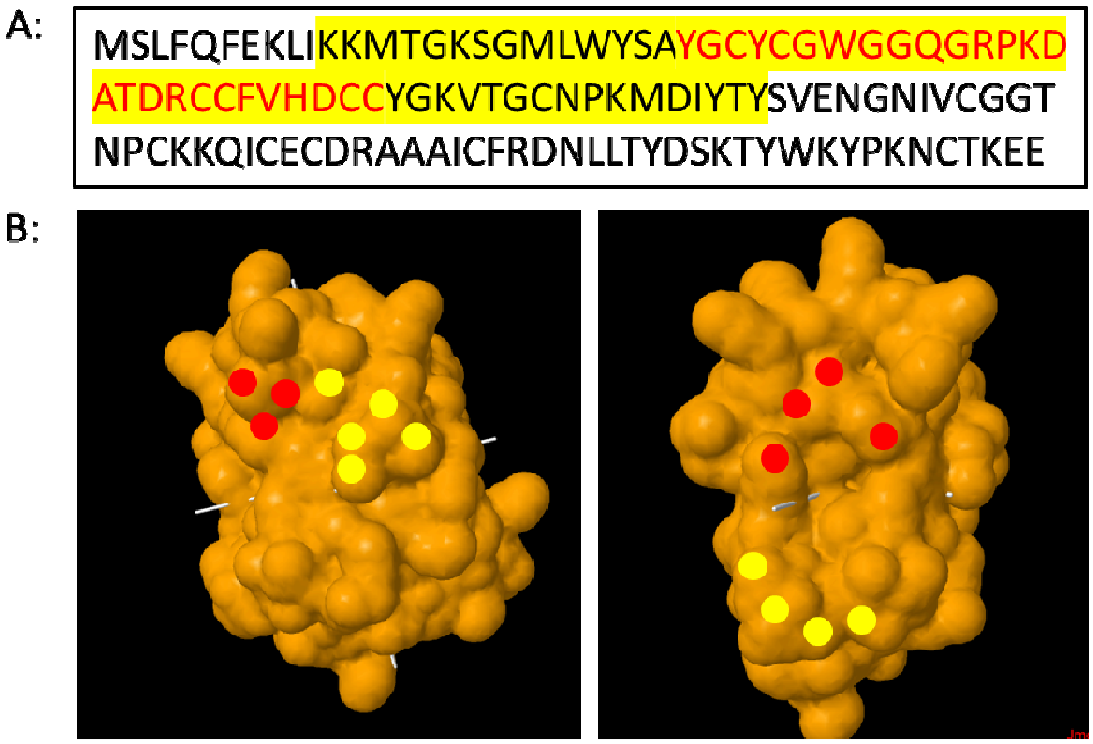
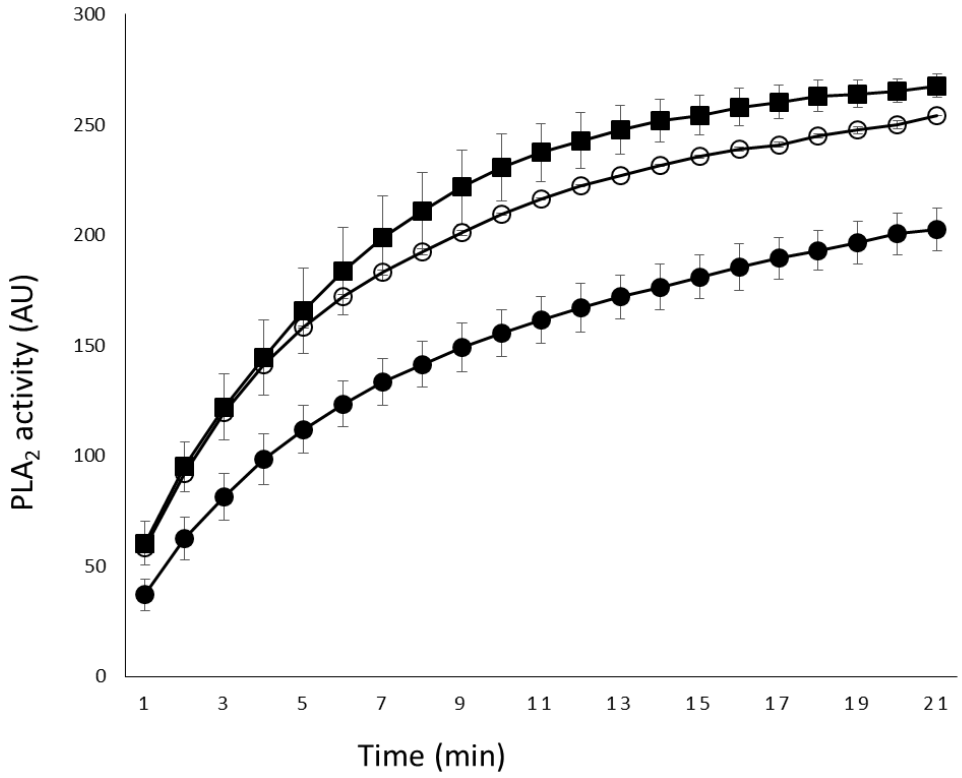


Figure 1. *In silico* design of consensus peptide from Cottonmouth PLA_2. **A.** Homology sequence of PLA_2 from cottonmouth species. This sequence corresponds to the homologous sequence between three PLA_2 proteins from cottonmouth species. The yellow section corresponds to the active site and was synthesized as the target molecule for panning. Characters in red are the metal binding site, based on BLAST annotations. **B.** Crystal Structure of *A. p. leucostoma* PLA_2. The front and 90° counterclockwise rotated views of PLA_2 are shown. Colored circles annotate shared solvent-exposed residues with the synthesized homologous sequence (red: metal binding site, yellow: active site).



375

376 **Figure 2.** Anti-PLA_2 activity of polyclonal M13 phages after the primary panning. Two different

377 concentration of polyclonal anti-PLA_2 M13 phages after the primary panning inhibited 55% and

378 15% of PLA_2 activity of Western cottonmouth venom after 30min incubation. [■: venom only

379 (4.88µg/ml); ○: venom with $8x10^{11}$ PFU/ml of anti-PLA_2 phages; ●: venom with $3.2x10^{12}$

380 PFU/ml of anti-PLA_2 phages.]

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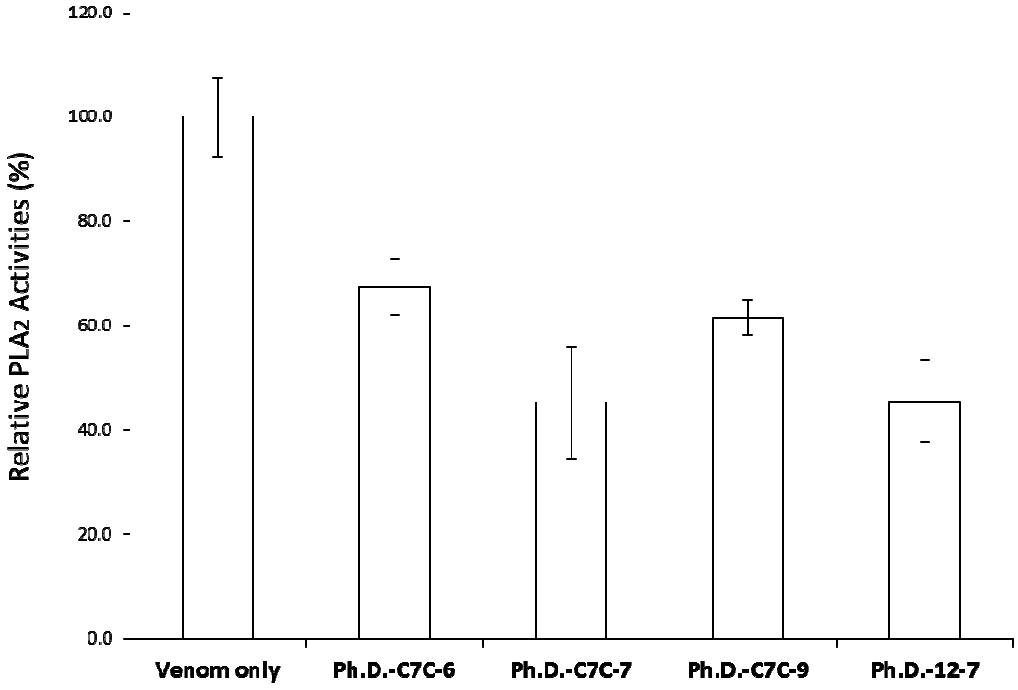


382

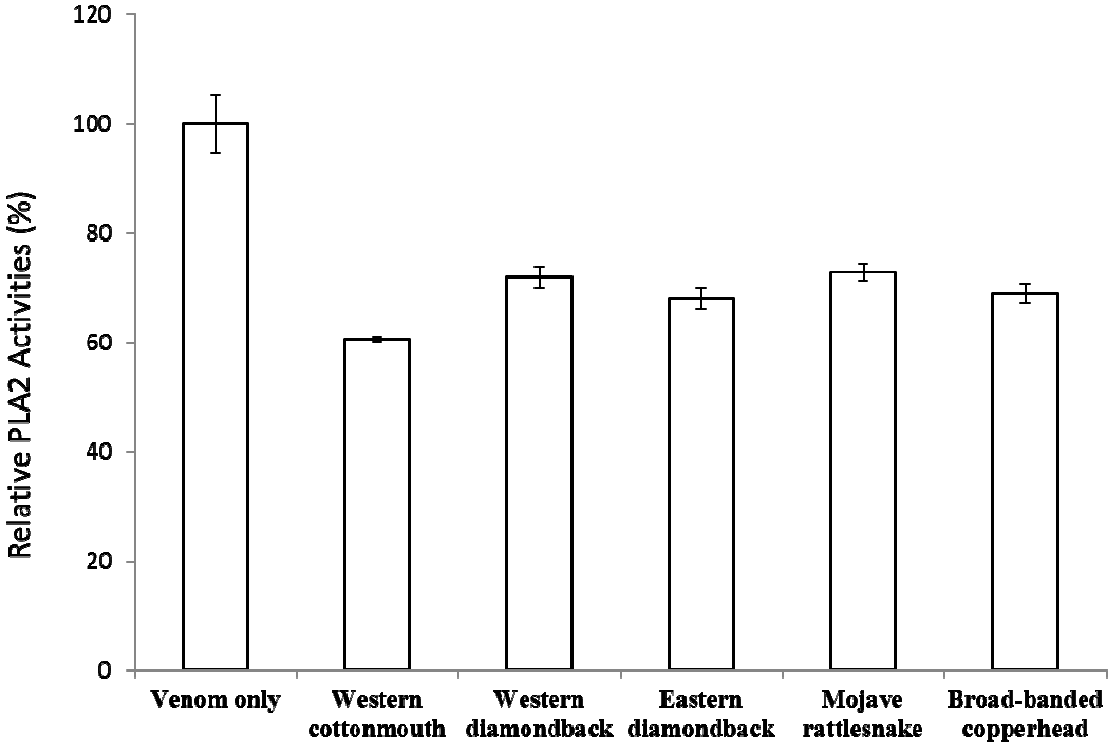
383 **Figure 3.** Anti-PLA_2 activities of selected M13 phage clones. Incubation with selected phage

384 clones isolated from phage display libraries inhibited the PLA_2 activity of *A. p. leucostoma*

385 venom by 30-60% during the first 10min. Phage clones were incubated with 4.88µg/ml of venom

386 for 30min.

387



388

389 **Figure 4.** Cross-species anti-PLA_2 activity of Ph.D.-12-7 phages against five major snake
390 venoms in North America. Cross-species anti-PLA_2 activity was tested against five major snake
391 venoms in North America (Western cottonmouth, Eastern diamondback rattlesnake, Western
392 diamondback rattlesnake, Mojave rattlesnake, and broad-banded copperhead) using one of
393 selected anti-PLA_2 clones (Ph.D.-12-7). Approximately 40% inhibition in Western cottonmouth
394 venom and 30% inhibition in the other crotalid venoms were observed.

395

Table 1: Nucleotide sequences and corresponding peptide sequences of selected monoclonal phage binding motifs.

Monoclonal Isolate	Nucleotide Sequence	Peptide Sequence
Ph.D.-C7C-6	TCGCCGTTGCATAAGACTATG	SPLHKTM
Ph.D.-C7C-7	TCGGGGATGAAGAAGACGAAG	SGMKKTK
Ph.D.-C7C-9	AAGACGACGAAGATGGGGTTG	KTTKMGL
Ph.D.-12-7	AAGCTTATTCATGGTAATGGTGTTATGGATGAGGGG	KLIHGNGVMDEG