

1 **MINI-REVIEW**

2
3 **Ticks and the effects of their saliva on growth factors involved in skin**
4 **wound healing**

5
6 Pavlína Bartíková^{1,*}, Mária Kazimírová² and Iveta Štibrániová¹

7
8 ¹Biomedical Research Center, Institute of Virology, Slovak Academy of Sciences, Dúbravská cesta 9, 845 05
9 Bratislava, Slovakia

10 ²Institute of Zoology, Slovak Academy of Sciences, Dúbravská cesta 9, 845 06 Bratislava, Slovakia

11
12 ***Correspondence to:** Pavlína Bartíková, Email: virupaca@savba.sk

13
14 **Received:** 10 June 2020 | **Revised:** 18 September 2020 | **Accepted:** 22 September 2020 | **Published:** 22 September
15 2020

16
17 *J Venom Res* (2020), Vol 10, 00-00

18
19 **© Copyright** The Author(s). This is an open access article, published under the terms of the Creative Commons
20 Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/4.0>). This license permits non-
21 commercial use, distribution and reproduction of this article, provided the original work is appropriately
22 acknowledged, with correct citation details.

23
24 **Note:** *This is not the final version of this article, which will be available in the near future.*

26 **ABSTRACT**

27
28 Ticks are unique hematophagous arthropods and possess an astounding array of salivary molecules that
29 ensure their unnoticed and prolonged attachment to the host skin. Furthermore, ticks are very effective
30 vectors of a diverse spectrum of pathogens. In order to feed, tick chelicerae cut the host epidermis and
31 their hypostome penetrates through the layers of the skin. As a result of laceration of the skin and
32 rupturing blood vessels, a pool of blood is formed in the dermis, serving for intermittent blood sucking
33 and secretion of saliva. Cutaneous injury caused by tick mouthparts should normally elicit wound
34 healing, a complex biological process coordinated by interaction among different host cells, numerous
35 signalling pathways and by a variety of soluble factors including growth factors. Growth factors,
36 endogenous signalling proteins involved in various biological events, are key players in all phases of the
37 skin repair process. Maintaining feeding site integrity by overcoming sequential phases of wound
38 healing is particularly important for ixodid ticks and is governed by bioactive molecules in their saliva.
39 Tick saliva is a complex mixture of proteins, peptides, and non-peptide molecules and its composition
40 depends on the feeding phase, tick developmental stage, gender and/or the presence/absence of
41 microbial agents. In addition to already demonstrated anti-haemostatic, anti-cytokine and anti-
42 chemokine activities, anti-growth factors activities were also detected in saliva of some tick species. In
43 consequence of counteracting host defences by ticks, tick-borne pathogens can be transmitted to and
44 disseminated in the host. Elucidation of the complex interplay between ticks – pathogens – host
45 cutaneous immunity could lead to improved vector and pathogens control strategies. Additionally, tick
46 saliva bioactive molecules have a promising therapeutic perspective to cure some human diseases
47 associated with dysregulation of specific cytokines/growth factors and alterations in their signalling
48 pathways.

49

50 **KEYWORDS:** Ticks, saliva, growth factors, wound healing, host immunity

51

52 **INTRODUCTION**

53

54 Ticks are hematophagous ectoparasites with immense medical and veterinary importance due to
55 transmission of a wide spectrum of pathogens and direct damage of host skin. However, based on
56 discovering toxins in tick saliva that are similar to those found in venomous animals, such as spiders or
57 snakes, ticks should also be referred to as venomous blood-feeding arthropods (Cabezas-Cruz and
58 Valdés, 2014). To successfully obtain a blood meal, ticks as telmophages lacerate all epidermis layers by
59 penetration of chelicerae and hypostome and, by rupturing blood vessels, they create a blood pool in
60 the dermis. A continuous feeding process till repletion, lasting for several hours to days, is boosted and

61 ensured by tick salivary bioactive molecules. Tick saliva represents a very complex fluid secreted by
62 salivary glands into the bite site and contains an incredible number and variety of proteins, peptides and
63 non-peptide molecules that: i) facilitate the feeding process by overcoming haemostasis and wound
64 healing processes; ii) interfere with host innate and adaptive immune defences; ii) support transmission
65 and local establishment of pathogens in the immunomodulated feeding site (reviewed in Kotál et al,
66 2015; Kazimírová et al; 2017; Mans et al, 2017; Šimo et al, 2017; Nuttall, 2018; Wikel, 2018); iii) can
67 induce paralysis or other toxicoses in vertebrate hosts (Cabezas-Cruz and Valdés, 2014; Pienaar et al,
68 2018).

69
70 Skin injury elicited by tick feeding initiates immediately a repair process which is composed of several
71 stages generally grouped into sequential overlapping phases: inflammation, proliferation and tissue
72 remodelling (Canedo-Dorantes and Canedo-Ayla, 2019). Each phase is conducted by a complex interplay
73 among several resident skin cells including keratinocytes, fibroblasts, endothelial cells, macrophages,
74 platelets and infiltrating immune cells and is performed and controlled by an equally intricate signalling
75 network of numerous cytokines, chemokines and growth factors (Sorg et al, 2017; Larouche et al, 2018).

76
77 Growth factors (GFs), naturally occurring signalling molecules, are involved in infinite biological events
78 including cell growth, proliferation, migration and differentiation, and are irreplaceable in the wound
79 healing process. GFs play critical roles in modulating inflammatory responses, enhancing granulation
80 tissue formation, and in inducing angiogenesis. They are essential for successful extracellular matrix
81 (ECM) formation and tissue remodelling processes (Park et al, 2017).

82
83 Despite the undoubted key role of GFs in wound healing, data about interactions between ticks and GFs
84 are very scarce. This mini-review contains information about effects of tick salivary molecules on
85 vertebrate GFs, key players of wound healing, and discusses their possible therapeutic potential.

86

87 **SKIN**

88

89 The interaction between ticks, tick-borne pathogens and the host immune response is a complex and
90 multifactorial event taking place in the vertebrate host skin. The skin is no longer considered just as a
91 mechanical barrier, but represents a sophisticated intricate communicative line between inner and
92 external environment of individuals. The discoveries of the 'skin immune system', 'keratinocytes as
93 potent sensors of infectious intruders and danger signals' and more recently of 'the skin microbiome'
94 highlighted the skin as complex multitasking organ with unique immunologic properties (DiMeglio et al,
95 2011). Under constant exposure to environmental stimuli, the skin performs numerous tasks to preserve

96 homeostasis and integrity of the organism. This ability of multifunctionality is closely linked to the skin
97 anatomical structure. Skin consists of three major layers – epidermis, dermis and hypodermis. The
98 epidermis, a four layered stratified epithelium, houses mostly keratinocytes (90%), melanocytes and
99 immune cells, such as Langerhans cells and dendritic epidermal T-lymphocytes. The composition of
100 dermis is more varied with greater cell diversity (fibroblasts, dendritic cells, macrophages, natural killer
101 cells, mast cells, T-lymphocytes) and thin and thick collagen fibres, respectively. Close to a basement
102 membrane, there is a papillary layer of dermis with high fibroblast density and thin collagen fibres
103 followed by reticular dermis consisting of thick collagen fibres and low cell density. Hypodermis, the
104 white dermal adipose tissue, consists of pre- and mature adipocytes surrounded by fibroblasts, nerves
105 and blood vessels (Pasparakis et al, 2014; Rognoni and Watt, 2018).

106

107 Despite serving as a significant and effective barrier, the skin is still the major entry portal for most
108 pathogens transmitted by arthropod vectors such as ticks, mosquitoes, sand flies, etc. during their
109 feeding, taking advantage of their ability to penetrate skin and bypass the host defence responses in
110 wounds evoked by mechanical injury (Boulanger, 2018).

111

112 **WOUND HEALING**

113

114 As a response to damage or disturbance of the skin's barrier function, wound healing is initiated. Wound
115 healing is an evolutionarily conserved, complex, multicellular process that, in skin, aims to protect and
116 restore tissue integrity. Formation of new tissue and wound closure involves the coordinated efforts of
117 myriad cell types including resident skin cells as well as infiltrated immunocompetent cells and is
118 executed and regulated by a complex signalling network involving numerous GFs, cytokines and
119 chemokines (Behm et al, 2012; Rodrigues et al, 2019). Sequential overlapping, and functionally
120 coordinated phases of acute cutaneous wound healing are activated within seconds of injury and evolve
121 over days, weeks, and months until skin integrity is restored (Greaves et al, 2013; Eming et al, 2014).

122

123 **HAEMOSTASIS/INFLAMMATION PHASE**

124

125 Upon the tick hypostome penetration into the skin, a clotting cascade is initiated with the aid of
126 activated platelets, resulting in the formation of a coagulation fibrin clot and ensures haemostasis. The
127 fibrin clot also provides a basic matrix to support inflammatory (neutrophils and monocytes) or other
128 resident skin cells (fibroblasts, endothelial cells) influx. Activated platelets, trapped in the clot, express
129 adhesion receptors for interaction with fibroblasts and endothelial cells and for recruitment of
130 circulatory cells and simultaneously release various GFs (Figure 1) and chemokines to regulate all

131 downstream phase events of wound healing (Eming et al, 2014; Golebiewska and Poole, 2015). Of GFs,
132 platelets secrete platelet-derived growth factor (PDGF), transforming growth factor- α (TGF- α),
133 transforming growth factor- β (TGF- β), epidermal growth factor (EGF), heparin-binding EGF (HP-EGF),
134 insulin-like growth factor 1 (IGF-1), vascular endothelial growth factor (VEGF), and fibroblast growth
135 factor (FGF) that interact, promote and chemotactically attract cells into the wound. PDGF attracts
136 neutrophils to the wound; VEGF and FGF endothelial cells and initiate angiogenesis.

137
138 Along with haemostasis, early inflammatory reactions trigger innate defensive reactions at the wound
139 site, specifically neutrophils, the first immune cells entering the wound followed by monocytes, which
140 convert into macrophages by TGF- β . Both neutrophils and macrophages control and kill invading
141 pathogens and amplify inflammatory response by secretion of antimicrobial molecules, cytokines and
142 GFs. Production of FGF, EGF, TGF- β , VEGF and PDGF leads to tissue granulation and transition to the
143 proliferative phase (Greaves et al, 2013).

144

145 **PROLIFERATIVE PHASE AND REMODELLING PHASE (MATURATION)**

146

147 The proliferative phase involves numerous important cellular and molecular components that
148 contribute to granulation tissue formation and ECM and initiation of angiogenesis. The proliferative
149 phase is characterized by: i) fibroplasia (proliferation of fibroblasts and their differentiation to
150 myofibroblasts, deposition of EMC) driven by TGF- β 1, EGF, FGF-2, PDGF and cytokines; ii) re-
151 epithelization (differentiation, replication and migration of keratinocytes) stimulated by EGF, hepatocyte
152 growth factor (HGF), FGFs, IGF-1, TGF- β 1; and iii) angiogenesis coordinated by VEGF, VEGF-A, FGF-2,
153 PDGF, TGF- β 1 (Greaves et al, 2013; Canedo-Dorantes and Canedo-Ayla, 2019; Rousselle et al, 2019).
154 Restoration of damaged blood vessels and growth of new ones is an essential part of healing, as it
155 provides nutrition and oxygen to the cells in the wound. Finally, a transition from granulation tissue to
156 mature scar occurs, characterized by continued collagen synthesis and collagen catabolism. The
157 remodelling phase is characterized by the active reorganization of the ECM, a reduction in the total
158 number of capillaries and replacement of type III collagen by type I collagen.

159

160 According to above mentioned studies on the wound healing process it is evident that besides the
161 importance of cell-cell and cell-matrix interactions, different GFs are involved and play critical roles in all
162 stages of the repair process.

163

164 **GROWTH FACTORS INVOLVED IN WOUND HEALING**

165

166 GFs comprise a large group of regulatory proteins affecting cellular growth, proliferation, differentiation,
167 and/or migration. They can be secreted as fully functional molecules or as molecules that require
168 further posttranslational processing in order to be activated. These chemical messengers attach to
169 specific cell surface receptors and via this interaction mediate inter and intracellular signalling pathways
170 by paracrine, autocrine, juxtacrine, or endocrine mechanisms (Barrientos et al, 2008; Fabi and
171 Sundaram, 2014). GFs are important in both normal physiological processes such as wound healing and
172 abnormal processes such as cancer. In the skin wound healing, GFs synthesized by fibroblasts,
173 keratinocytes, platelets, lymphocytes, and mast cells play critical roles in modulating inflammatory
174 responses, enhancing granulation tissue and ECM formation, inducing angiogenesis and tissue
175 remodelling (Barrientos et al, 2008; Behm et al, 2012). Supplementary Table S1 shows cell sources, cell
176 targets and some mechanisms of action that have been identified for the key GFs involved in wound
177 healing. The expression and function of GFs in normal skin wound healing correlate spatially and
178 temporally with the phase-specific recruitment of different cells.

179
180 However, GFs deficiencies, including reduced levels, are responsible for chronic non-healing wounds. On
181 the other hand, abnormal GFs expression is associated with impaired wound healing or excessive
182 scarring (fibrosis) (Lichtman et al, 2016; Park et al, 2017).

183

184 **TICK COUNTERMEASURES AGAINST GROWTH FACTORS**

185

186 All blood-feeding arthropods, especially ticks, face challenges from host haemostasis, inflammation, and
187 initiation of immune responses that could impair subsequent blood feeding. The feeding process of hard
188 ticks is a sequence of behavioural changes from questing and engaging the host, finding a suitable
189 feeding site, penetration of the skin, blood pool creation and feeding to repletion. Thus, host resident
190 skin cells are exposed to both, mechanical injury from insertion of mouthparts and biological actions of
191 saliva molecules facilitating blood meal acquisition. To secure uninterrupted blood uptake, ticks
192 suppress and evade the complex physiological host immune and homeostatic responses that are raised
193 against them. Cutaneous injury and ruptured blood vessels should normally elicit a wound healing.

194 Repairing progression would cause tick rejection and/or disrupt tick feeding, however, ticks evolved the
195 ability to overcome all host defence responses, including wound healing so as to maintain the feeding
196 lesion in dermis and complete the feeding process. Tick salivary glands produce and secrete a cocktail of
197 different molecules (Figure 1) that interfere with various components of haemostasis, wound healing,
198 and both arms of the immune system of the vertebrate hosts, including enzymes, cytokines,
199 complement and even GFs (Kazimírová and Štibrániová, 2013; Wikel, 2018; Štibrániová et al, 2019).
200 Studies of Hajnicka et al (2011) and Slovák et al (2014) determined a direct impact of salivary gland

201 extracts (SGE) derived from different hard tick species on several GFs involved in wound healing. They
202 discovered various anti-TGF- β 1, -PDGF, -HGF and -FGF-2 activities depending on the tick species (see
203 Table 1). Specific binding activity was confirmed just for TGF- β 1. No anti-EGF, -granulocyte macrophage
204 colony stimulating factor (GM-CSF), -keratinocyte growth factor (KGF) and -VEGF activities have been
205 demonstrated in any of the tick SGE. Of all tested ticks, SGE of *Amblyomma variegatum* and *Hyalomma*
206 *excavatum* showed a richer repertoire of GF-binding molecules than the other tested species. In
207 addition to anti-GF effects, SGE derived from ticks with long mouthparts displayed *in vitro* anti-
208 proliferative and cytotoxic activities associated with actin filaments disruption in different cell lines. The
209 anti-GFs activities identified by Hajnická et al (2011) could be one of the explanations of suppressive
210 effects of *Dermacentor variabilis* SGE and saliva on fibroblast migration and ability to repair an injury
211 that were previously observed by Kramer et al (2008). Tick SGE/saliva also reduced pro-migration
212 signalling triggered with extracellular signal-regulated kinase (ERK) in PDGF-stimulated fibroblasts. On
213 the other side, *D. variabilis* saliva enhanced basal- and PDGF-stimulated migration of macrophage-
214 derived IC-21 cell line without affecting ERK signalling and suppressed macrophage ability to
215 phagocytose zymosan particles (Kramer et al, 2011).

216
217 The ERK signalling pathway, like other MAPK serine/ threonine kinases pathways (JNK, p38MAPK),
218 regulates cell migration induced by GFs, including PDGF, VEGF, EGF, FGF and insulin. Thus, ticks could
219 conquer wound healing by inhibiting the signalling activity as shown earlier. In addition, SGE of *D.*
220 *variabilis* was found to display the same effects on renal epithelial OK cells (not concerned in wound
221 healing) suggesting that tick saliva might be valuable in controlling the migration of many cell types,
222 including endothelial cells during angiogenesis and invasiveness of cancer cells (Kramer et al, 2008).
223 Studies that focused on beneficial effects of tick saliva and their therapeutic potential are mentioned
224 below.

225
226 In *Amblyomma americanum* saliva three insulin-like GFs binding-related proteins (IGFBP-rPs) were
227 identified and termed as AamIGFBP-rP1 and AamIGFBP-rP6S (short) and AamIGFBP-rP6L (long),
228 respectively, according to comparison with human IGFBP-rPs. IGFBP-rPs, belonging to the IGF system,
229 are responsible for cell differentiation and growth in vertebrates. The RNAi-mediated silencing of
230 AamIGFBP-rPs suggested that these three proteins may be involved in physiological functions linked to
231 the tick feeding process, perhaps through possible AamIGFBP-rP1 stimulation of prostacyclin secretion
232 from salivary glands into the tick saliva (Mulenga and Khumthong, 2010). Both AamIGFBP-rP6 are highly
233 expressed during the first 24 h of feeding and are probably associated with formation of the feeding
234 site, but are downregulated as ticks continue to feed. AamIGFBP-rP1, which is upregulated after feeding
235 commences, may be associated with regulating continuous events such as blood ingestion and/or

236 maintenance of the tick-feeding lesion. A study with recombinant AamIGFBP-rP1 revealed its ability to
237 bind insulin, but no IGFs, and showed no antihaemostatic or antimicrobial functions (Radulović et al,
238 2015). Native AamIGFBP-rP1 protein is an immunogen and, as previously observed, conservation of
239 AamIGFBP-rPs amino acid sequences amongst other hard tick species makes them potential targets for
240 universal anti-tick vaccine development (Mulenga and Khumthong, 2010; Radulović et al, 2015). Bakshi
241 et al (2019) showed that rAamIGFBP-rPs belong to the pro-inflammatory *A. americanum* saliva proteins
242 or, in other words, pro-host defence proteins, which stimulate both PBMC-derived and mouse RAW
243 267.4 macrophages to *in vitro* express pro-inflammatory markers such as interleukin (IL)-1, IL-6 and
244 tumour necrosis factor (TNF)- α . On the contrary, anti-inflammatory or anti-host defence tick saliva
245 proteins, such as the two serpins derived from *A. americanum*, were found to enhance expression of
246 anti-inflammatory cytokines (IL-10 and TGF- β) in the macrophages pre-activated by LPS or rAamIGFBP-
247 rPs. Mice paw oedema test confirmed *in vivo* functional countervailing of tick saliva pro- and anti-
248 inflammatory proteins (Bakshi et al, 2019).

249
250 Despite having much shorter feeding times, argasid (soft) ticks may manipulate host wound healing
251 responses as well. *Ornithodoros brasiliensis*, a nidicolous soft tick, is frequently allied with human and
252 canine toxicosis syndrome up to severe reactions and with notably sluggish healing lesion at the bite
253 site, which can take several weeks to heal. Saliva and/or salivary gland homogenates from *O. brasiliensis*
254 ticks significantly delayed *in vivo* wound healing of rat skin in the classical model of excisional induced
255 skin lesions and showed a strong cytotoxic and antiproliferative activity on cultured endothelial cells *in*
256 *vitro*. However, the mechanisms of the described effects are unknown (Reck et al, 2013).

257

258 THERAPEUTIC POTENTIAL OF TICK SALIVA

259

260 Disregarding the parasitic life style and transmission of pathogens, the ability of ticks to defeat every
261 stage of wound healing and host defences by a wide array of saliva constituents shift them to a spotlight
262 as promising sources of new drugs (Sousa et al, 2015; Chmelar et al, 2019; Štibraniova et al, 2019).
263 Among the various tick saliva activities, anti-angiogenic properties and also cytotoxic effects on various
264 cell types were observed. Angiogenesis is a crucial event involved in many physiological processes from
265 early embryonic to adult stages including the recovery of haemostasis during wound repair (Wietecha
266 and Dipietro, 2013). Yet, in some severe pathological processes including cancer (Welti et al, 2013),
267 angiogenesis has become a target for treatment. Of GFs, VEGF-A is the most important mediator of
268 angiogenesis. Amblyomin-X, a 15 kDa recombinant Kunitz-type serine protease inhibitor of coagulation
269 factor Xa (FXa) derived from *Amblyomma cajennense* ticks (Batista et al, 2010) was found to possess
270 anti-tumour and anti-metastatic activities (Chudzinski-Tavassi et al, 2010; Ventura et al, 2013) and also

271 anti-VEGF-A activity. Amblyomin-X treatment did not affect VEGF-A induced cell migration, but inhibited
272 VEGF-A induced angiogenesis in the mouse dorsal subcutaneous tissue as well as in the chicken
273 chorioallantoic membrane. Amblyomin-X also reduced cell adhesion and tube formation and inhibited
274 VEGF-A induced endothelial PECAM-1 expression (Drewes et al, 2012).

275
276 Another potent anti-angiogenesis factor with a potential to control cancer is the Troponin I-like molecule
277 (HLTnI) identified in saliva of *Haemaphysalis longicornis* by You et al (2001). Recombinant HLTnI significantly
278 suppressed the capillary development in human vascular endothelial cells (HUVEC) in a dose-dependent
279 manner in VEGF competitive angiogenesis assay *in vitro* (Fukumuto et al, 2006). The mechanism of this effect
280 is unclear, however, the structure similarity with human troponin I suggests an interaction of HLTnI with
281 receptors for VEGF and FGF-2 on endothelial cells. On the other hand, HLTnI can facilitate blood feeding of
282 ticks by inhibition of angiogenesis and moreover, it can be a potent target molecule to control ticks due to its
283 strong immunogenicity.

284
285 Ixolaris, an effective Kunitz domain anticoagulant derived from salivary glands of *Ixodes scapularis* targeting
286 the FVIIa/TF complex (Francischetti et al, 2002), displays promising anti-tumour therapeutic usage. Ixolaris was
287 found to decrease U87-MG tumour growth *in vivo* in a xenograft model in nude mice and also reduce tumour
288 vascularization and expression of VEGF (Carneiro-Lobo et al, 2009, 2012).

289

290 **SIGNALLING PATHWAYS**

291

292 Protein Kinase B (PKB)/Akt and extracellular signal-regulated kinase (ERK) signalling pathways can both
293 mediate GF-induced cellular migration, and aberrant regulation of Akt and ERK pathways can contribute to
294 cancer phenotypes (Gan et al, 2010; Liu et al, 2012). The MEK/ERK pathway, for example, is associated with
295 more than 30% of human cancers (Bonjardim, 2017).

296

297 Saliva from *D. variabilis* was observed to inhibit basal and agonist-induced Saos-2 and MB-231 migration and
298 invasion. Saliva of this tick species had no effect on EGF-stimulated Akt and ERK activity in MB-231 cells. In the
299 Saos-2 cells, saliva suppressed EGF-activation of Akt, however, only basal ERK activity was affected in MB-231
300 cells (Poole et al, 2013).

301

302 TGF- β 1, via specific serine/threonine kinase receptors, activates the common SMAD- dependent signalling
303 pathway (Massagué, 2012; Fabregat et al, 2014) and/or non-SMAD pathways regulated by ERK, p38 MAPK,
304 JNK, and PKB/AKT (Walton et al, 2012). The dysregulation of the TGF- β 1 pathways leads to numerous human
305 diseases and disorders, including cancer, fibrosis and inflammation (Pickup et al, 2013). SGE derived from

306 *Dermacentor reticulatus* and *Hyalomma anatolicum excavatum* ticks was observed to effectively decrease
307 human (hu)TGF- β 1-induced signalling pathways, canonical as well non-canonical in SiHa and HeLa cervical
308 cancer cell lines, varying in their response to huTGF- β 1. Furthermore, SGE of *D. reticulatus* significantly
309 inhibited huTGF- β 1-induced ERK1/2 activation, but not Akt activation. On the other hand, SGE of *H.*
310 *anatolicum excavatum* ticks did not display any evident effect on ERK1/2 or AKT activation induced by huTGF-
311 β 1 (Holikova et al, 2018). The results suggest that the anti-GF activities of SGE depend not only on the origin of
312 the treated cells but also on the tick species from which the SGEs originate. Additionally, crosstalk between
313 signalling pathways of different GFs has been described and isolation and identification of specific tick
314 molecules responsible for anti-GFs activities is highly desirable.

315

316 **CONCLUSIONS**

317

318 GFs and their signalling pathways are considered as crucial players in diverse physiological processes,
319 including wound healing. However, any aberration in these pathways is associated with many
320 immunological disorders and diseases, even cancer. Tick saliva comprises a complex cocktail of bioactive
321 compounds that facilitate their feeding process and transmission of pathogens. Saliva of some tick
322 species showed anti-GFs activities, but the knowledge on the mechanisms of action of tick salivary
323 molecules on GFs involved in wound healing in vertebrate skin is still limited. However, tick saliva
324 molecules with immunomodulatory, anti-inflammatory, anti-clotting, anti-platelet, anti-cytokine, anti-
325 GFs as well as anti-tumour and anti-angiogenic properties represent important sources for discovering
326 and designing new therapeutics for various pathways of the mammalian physiology.

327

328 **ACKNOWLEDGMENTS**

329

330 This work was supported by the Grant agency of Slovak Republic (VEGA 2/0047/18 and VEGA
331 2/0172/19). The authors thank Mrs Galina Jarabkova for her assistance with the designing of Figure 1 of
332 this article.

333

334 **COMPETING INTERESTS**

335

336 None declared.

337

338 **ABBREVIATIONS**

339

340 GFs: Growth factors

341 ECM: Extracellular matrix
342 EGF: Epidermal growth factor
343 FGF: Fibroblast growth factor
344 HP-EGF: Heparin-binding EGF
345 HGF: Hepatocyte growth factor
346 IGF-1: Insulin-like growth factor-1
347 PDGF: Platelet-derived growth factor
348 TGF- α Transforming growth factor-alpha
349 TGF- β : Transforming growth factor-beta
350 VEGF: Vascular endothelial growth factor
351 GM-CSF: Granulocyte macrophage colony stimulating factor
352 KGF: Keratinocyte growth factor
353 SGE: Salivary gland extract
354 ERK: Extracellular signal-regulated kinase

355

356 REFERENCES

357

- 358 Bakshi M, Kim TK, Porter L, Mwangi W and Mulenga A. 2019. *Amblyomma americanum* ticks utilizes
359 countervailing pro and anti-inflammatory proteins to evade host defense. *PLoS Pathog*, 15, e1008128.
- 360 Barrientos S, Stojadinovic O, Golinko MS, Brem H and Tomic-Canic M. 2008. Growth factors and
361 cytokines in wound healing. *Wound Repair Regen*, 16, 585-601.
- 362 Batista IF, Ramos OH, Ventura JS, Junqueira-de-Azevedo IL, Ho PL and Chudzinski-Tavassi AM. 2010. A
363 new Factor Xa inhibitor from *Amblyomma cajennense* with a unique domain composition. *Arch Biochem*
364 *Biophys*, 493, 151-156.
- 365 Behm B, Babilas P, Landthaler M and Schreml S. 2012. Cytokines, chemokines and growth factors in
366 wound healing. *J Eur Acad Dermatol Venereol*, 26, 812-820.
- 367 Bonjardim CA. 2017. Viral exploitation of the MEK/ERK pathway – A tale of vaccinia virus and other
368 viruses. *Virology*, 507, 267-275.
- 369 Boulanger N (Ed). 2018. *Skin and Arthropod Vectors*. Elsevier Inc, London, UK.
- 370 Cabezas-Cruz A and Valdés JJ. 2014. Are ticks venomous animals? *Front Zool*, 11, 1-18.
- 371 Cañedo-Dorantes L and Cañedo-Ayala M. 2019. Skin acute wound healing: A comprehensive review. *Int J*
372 *Inflam*, 2019, Article ID 3706315.
- 373 Carneiro-Lobo TC, Konig S, Machado DE, et al. 2009. Ixolaris, a tissue factor inhibitor, blocks primary
374 tumor growth and angiogenesis in a glioblastoma model. *J Thromb Haemost*, 7, 1855-1864.

375 Carneiro-Lobo TC, Schaffner F, Disse J, et al. 2012. The tick-derived inhibitor Ixolaris prevents tissue
376 factor signaling on tumor cells. *J Thromb Haemost*, 10, 1849-1858.

377 Chmelař J, Kotál J, Kovaříková A and Kotsyfakis M. 2019. The use of tick salivary proteins as novel
378 therapeutics. *Front Physiol*, 10, 1-10.

379 Chudzinski-Tavassi AM, de-Sá-Júnior PL, Simons SM, et al. 2010. A new tick Kunitz type inhibitor,
380 Amblyomin-X, induces tumor cell death by modulating genes related to the cell cycle and targeting the
381 ubiquitin-proteasome system. *Toxicon*, 56, 1145-1154.

382 Di Meglio P, Perera GK and Nestle FO. 2011. The multitasking organ: Recent insights into skin immune
383 function. *Immunity*, 35, 857-869.

384 Drewes CC, Dias RYS, Hebeda CB, et al. 2012. Actions of the Kunitz-type serine protease inhibitor
385 Amblyomin-X on VEGF-A-induced angiogenesis. *Toxicon*, 60, 333-340.

386 Eming SA, Martin P and Tomic-Canic M. 2014. Wound repair and regeneration: Mechanisms, signaling,
387 and translation. *Sci Transl Med*, 6, 1-36.

388 Fabi S and Sundaram H. 2014. The potential of topical and injectable growth factors and cytokines for
389 skin rejuvenation. *Facial Plast Surg*, 30, 157-171.

390 Fabregat I, Fernando J, Mainez J and Sancho P. 2014. TGF-beta signaling in cancer treatment. *Curr*
391 *Pharm Design*, 20, 2934-2947.

392 Francischetti IMB, Valenzuela JG, Andersen JF, Mather TN and Ribeiro JMC. 2002. Ixolaris, a novel
393 recombinant tissue factor pathway inhibitor (TFPI) from the salivary gland of the tick, *Ixodes scapularis*:
394 identification of factor X and factor Xa as scaffolds for the inhibition of factor VIIa / tissue factor
395 complex. *Blood*, 99, 3602-3612.

396 Fukumoto S, Sakaguchi T, You M, Xuan X and Fujisaki K. 2006. Tick troponin I-like molecule is a potent
397 inhibitor for angiogenesis. *Microvasc Res*, 71, 218-221.

398 Gan Y, Shi C, Inge L, Hibner M, Balducci J and Huang Y. 2010. Differential roles of ERK and Akt pathways
399 in regulation of EGFR-mediated signaling and motility in prostate cancer cells. *Oncogene*, 29, 4947-4958.

400 Golebiewska EM and Poole AW. 2015. Platelet secretion: From haemostasis to wound healing and
401 beyond. *Blood Rev*, 29, 153-162.

402 Greaves NS, Ashcroft KJ, Baguneid M and Bayat A. 2013. Current understanding of molecular and
403 cellular mechanisms in fibroplasia and angiogenesis during acute wound healing. *J Dermatol Sci*, 72, 206-
404 217.

405 Hajnická V, Vančová-Štibrániová I, Slovák M, Kocáková P and Nuttall PA. 2011. Ixodid tick salivary gland
406 products target host wound healing growth factors. *Int J Parasitol*, 41, 213-223.

407 Holíková V, Štibrániová I, Bartíková P, Slovák M and Kazimírová M. 2018. Ixodid tick salivary gland
408 extracts suppress human transforming growth factor- β 1 triggered signalling pathways in cervical
409 carcinoma cells. *Biologia*, 73, 1109-1122.

410 Kazimírová M and Štibrániová I. 2013. Tick salivary compounds: their role in modulation of host
411 defences and pathogen transmission. *Front Cell Infect Microbiol*, 3, 43.

412 Kazimírová M, Thangamani S, Bartíková P, et al. 2017. Tick-borne viruses and biological processes at the
413 tick-host-virus interface. *Front Cell Infect Microbiol*, 7, 339.

414 Kotál J, Langhansová H, Lieskovská J, et al. 2015. Modulation of host immunity by tick saliva. *J*
415 *Proteomics*, 128, 58-68.

416 Kramer CD, Nahmias Z, Norman DD, Mulvihill TA, Coons LB and Cole JA. 2008. *Dermacentor variabilis*:
417 Regulation of fibroblast migration by tick salivary gland extract and saliva. *Exp Parasitol*, 119, 391-397.

418 Kramer CD, Poole NM, Coons LB and Cole JA. 2011. Tick saliva regulates migration, phagocytosis, and
419 gene expression in the macrophage-like cell line, IC-21. *Exp Parasitol*, 127, 665-671.

420 Larouche J, Sheoran S, Maruyama K and Martino MM. 2018. Immune regulation of skin wound healing:
421 Mechanisms and novel therapeutic targets. *Adv Wound Care*, 7, 209-231.

422 Lichtman MK, Otero-Vinas M and Falanga V. 2016. Transforming growth factor beta (TGF- β) isoforms in
423 wound healing and fibrosis. *Wound Repair Regen*, 24, 215-222.

424 Liu W, Wang K, Gong K, Li X and Luo K. 2012. Epidermal growth factor enhances MPC-38 pancreatic
425 cancer cell migration through upregulation of aquaporin3. *Mol Med Rep*, 6, 607-610.

426 Mans BJ, Featherston J, de Castro MH and Pienaar R. 2017. Gene duplication and protein evolution in
427 tick-host interactions. *Front Cell Infect Microbiol*, 7, 413.

428 Massagué J 2012. TGF β signalling in context. *Nat Rev Mol Cell Biol*, 13, 616-630.

429 Mulenga A and Khumthong R. 2010. Disrupting the *Amblyomma americanum* (L.) CD147 receptor
430 homolog prevents ticks from feeding to repletion and blocks spontaneous detachment of ticks from
431 their host. *Insect Biochem Mol Biol*, 40, 524-532.

432 Nuttall PA. 2018. Wonders of tick saliva. *Ticks Tick Borne Dis*, 10, 470-481.

433 Park JW, Hwang SR and Yoon I-S. 2017. Advanced growth factor delivery systems in wound management
434 and skin regeneration. *Molecules*, 22, 1259.

435 Pasparakis M, Haase I and Nestle FO. 2014. Mechanisms regulating skin immunity and inflammation. *Nat*
436 *Rev Immunol*, 14, 289-301.

437 Pickup M, Novitskiy S and Moses HL. 2013. The roles of TGF β in the tumour microenvironment. *Nat Rev*
438 *Cancer*, 13, 788-799.

439 Pienaar R, Neitz AWH and Mans BJ. 2018. Tick paralysis: Solving an enigma. *Vet Sci*, 5(2), 53.

440 Poole NM, Nyindodo-Ogari L, Kramer C, Coons LB and Cole JA. 2013. Effects of tick saliva on the
441 migratory and invasive activity of Saos-2 osteosarcoma and MDA-MB-231 breast cancer cells. *Ticks Tick*
442 *Borne Dis*, 4, 120-127.

443 Radulovic ZM, Porter LM, Kim TK, Bakshi M and Mulenga A. 2015. *Amblyomma americanum* tick saliva
444 insulin-like growth factor binding protein-related protein 1 binds insulin but not insulin-like growth
445 factors. *Insect Mol Biol*, 24, 539-550.

446 Reck J, Marks FS, Termignoni C, Guimarães JA and Martins JR. 2013. *Ornithodoros brasiliensis* (mouso
447 tick) salivary gland homogenates inhibit in vivo wound healing and in vitro endothelial cell proliferation.
448 *Parasitol Res*, 112, 1749-1753.

449 Rodrigues M, Kosaric N, Bonham CA and Gurtner GC. 2019. Wound healing: A cellular perspective.
450 *Physiol Rev*, 99, 665-706.

451 Rognoni E and Watt FM. 2018. Skin cell heterogeneity in development, wound healing, and cancer.
452 *Trends Cell Biol*, 28, 709-722.

453 Rousselle P, Braye F and Dayan G. 2019. Re-epithelialization of adult skin wounds: Cellular mechanisms
454 and therapeutic strategies. *Adv Drug Deliv Rev*, 146, 344-365.

455 Slovák M, Štibrániová I, Hajnická V and Nuttall PA. 2014. Antiplatelet-derived growth factor (PDGF)
456 activity in the saliva of ixodid ticks is linked with their long mouthparts. *Parasite Immunol*, 36, 32-42.

457 Sorg H, Tilkorn DJ, Hager S, Hauser J and Mirastschijski U. 2017. Skin wound healing: An update on the
458 current knowledge and concepts. *Eur Surg Res*, 58, 81-94.

459 Sousa ACP, Szabó MPJ, Oliveira CJF and Silva MJB. 2015. Exploring the anti-tumoral effects of tick saliva
460 and derived components. *Toxicon*, 102, 69-73.

461 Šimo L, Kazimirova M, Richardson J and Bonnet SI. 2017. The essential role of tick salivary glands and
462 saliva in tick feeding and pathogen transmission. *Front Cell Infect Microbiol*, 7, 281.

463 Štibrániová I, Bartíková P, Holíková V and Kazimírová M. 2019. Deciphering biological processes at the
464 tick-host interface opens new strategies for treatment of human diseases. *Front Physiol*, 10, 830.

465 Ventura JS, Faria F, Batista IF, et al. 2013. A Kunitz-type FXa inhibitor affects tumor progression,
466 hypercoagulable state and triggers apoptosis. *Biomed Pharmacother*, 67, 192-196.

467 Walton KL, Makanji Y and Harrison CA. 2012. New insights into the mechanisms of activin action and
468 inhibition. *Mol Cell Endocrinol*, 359, 2-12.

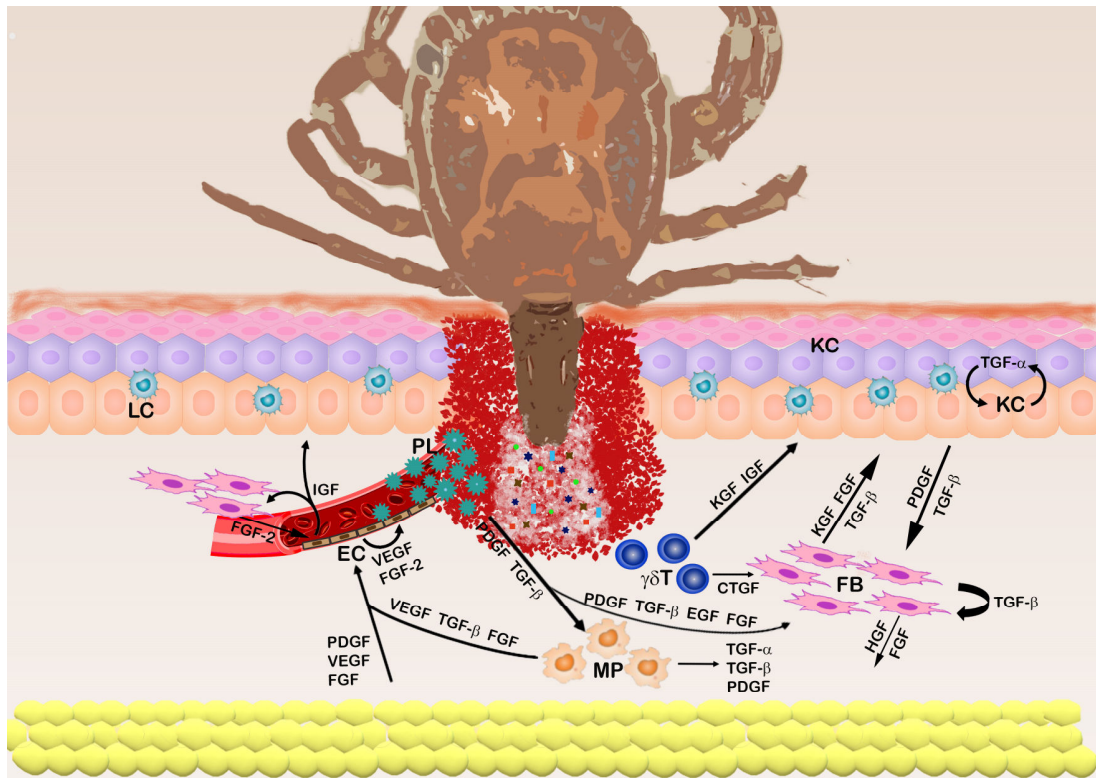
469 Welti J, Loges S, Dimmeler S and Carmeliet P. 2013. Recent molecular discoveries in angiogenesis and
470 antiangiogenic therapies in cancer. *J Clin Invest*, 123, 3190e3200.

471 Wietecha MS and Dipietro LA. 2013. Therapeutic approaches to the regulation of wound angiogenesis.
472 *Adv Wound Care*, 2, 81-86.

473 Wikel SK. 2018. Tick-host-pathogen systems immunobiology: an interactive trio. *Front Biosci*, 23, 265-
474 283.

475 You M, Xuan X, Tsuji N, et al. 2001. Molecular characterization of a troponin I-like protein from the hard
476 tick *Haemaphysalis longicornis*. *Insect Biochem Mol Biol*, 32, 67-73.

477



478
 479 **Figure 1.** Upon the tick hypostome penetration into the host skin, activated platelets, resident skin cells
 480 and infiltrated immunocompetent cells secrete numerous GFs into the wound edge. Platelets (PL) within
 481 the plug release PDGF, TGF- β , EGF, IGF, which are chemotactic for neutrophils, monocytes-macrophages
 482 and promote the proliferation of fibroblasts (FB), keratinocytes (KC) and endothelial cells (EC) in the
 483 wound. Due to a sophisticated network of sensors, KC recognize foreign or microbial agents and tissue
 484 damage and transmit warning by producing mediators (cytokines, chemokines, growth factors) and alert
 485 skin-resident immune cells, for instance, Langerhans cells (LC), type of dendritic cells resident in the
 486 epidermis representing the main antigen presenting cells. The $\gamma\delta$ T cells in dermis act as immune
 487 sentinels and moreover produce GFs important in wound healing, such as connective tissue growth
 488 factor (CTGF), FGF, KGF and IGF1. After injury, tissue-resident macrophages (MP) activate migration of
 489 circulating monocytes into the wound, which convert into MP by TGF- β . MP undergo phenotypic
 490 changes throughout the healing process, in the early stages they differentiate into the pro-inflammatory
 491 M1 subset of MP associated with phagocytic activity, scavenging as well as secrete pro-inflammatory
 492 cytokines and growth factors (FGF-2, PDGF, VEGF) to mobilize more immune cells and stimulate the
 493 proliferation of KC, FB and EC. During the proliferation stage of wound healing, M1 cells transform into a
 494 functionally and phenotypically anti-inflammatory M2 cells orchestrating the interaction with EC, FB and
 495 KC and GFs govern proliferation and differentiation of FB, KC and EC leading to tissue and vessels
 496 formation. Maintaining feeding site integrity by overcoming wound healing processes is particularly
 497 important for ixodid ticks and is governed by bioactive molecules in their saliva. Saliva of some tick
 498 species displayed anti-TGF- β , -PDGF, -FGF-2, -HGF, -VEGF activities.

499 **Table 1.** Anti-growth factor activities in salivary gland extracts of different tick species (modified
 500 according to Hajnická et al, 2011; Slovák et al, 2014).

Growth factors	<i>Amblyomma variegatum</i>		<i>Dermacentor reticulatus</i>		<i>Rhipicephalus appendiculatus</i>		<i>Hyalomma excavatum</i>			<i>Ixodes ricinus</i> <i>Ixodes scapularis</i>
	F	M	F	M	F	M	F	M	N	F
TGF-β1	++++	++++	++++	++++	++++	++	++++	++	++++	-
FGF-2	++++	++++	++	++	+	+	++++	++++	+	-
PDGF	++++	++++	-	-	-	-	++++	+	+++	++++
HGF	+++	++	+++	+	+++	++	++	++	++	-
EGF	-	-	-	-	-	-	nd	nd	nd	-
VEGF	-	-	-	-	-	-	nd	nd	nd	-
GM-CSF	-	-	-	-	-	-	nd	nd	nd	-
KGF	nd	nd	nd	nd	nd	nd	-	-	-	nd

501
 502 Activities are recorded as: nd (not done); - (undetectable); + (1–25%); ++ (26–50%); +++ (51–75%); ++++ (76–100%). Growth
 503 factor effects are shown as % reduction in OD reading compared with the control in specific ELISAs. F – adult females; M – adult
 504 males, N – nymphs